



SWAMI VIVEKANAND
SUBHARTI
UNIVERSITY
UGC Approved Meerut



AN ISO 21001: 2018 ORGANIZATION

OFFICE OF THE REGISTRAR

Gp Capt M Yakoob

M-in-D (Retd.), M.Tech.

REGISTRAR

registrar@subharti.org

Ref.No.U-508(i)/SVSU/2025/ 1622

Date:27.01.2025

NOTIFICATION

It is hereby notified for information of all the concerned that the Academic Council in its 34th meeting held on 25-07-2024 vide resolution No.34(9) has approved the revised syllabus (as per NEP-2020) of the following ordinance:

Ordinance No.V-135(B10), relating to B.Sc.-Biochemistry

The copy of above is enclosed and shall be applicable from Academic Session 2022-23 onwards.

This issues with the approval of the Hon'ble Vice Chancellor.

M Yakoob
27.01
2025
Registrar

Date: 27.01.2025

Ref.No.U-508(i)/SVSU/2025/ 1622

Copy forwarded to information of:

1. Hon'ble Vice-Chancellor
2. Controller of Examination
3. Dean-Academics
4. Director-IQAC
5. Dean-Faculty of Science (for compliance please)
6. CTO (with a request to upload the ordinance on University website)
7. Additional Registrar-Academics
8. Guard File

M Yakoob
27.01
2024
Registrar



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Subhartipuram, NH-58, Delhi-Haridwar Bypass Road, Meerut-250005 (U.P.) INDIA

Department of Biotechnology

Keral Verma Subharti College of Science

SWAMI VIVEKANAND SUBHARTI UNIVERSITY
MEERUT



Syllabus of B.Sc. Biochemistry (UG)
for Three Year Degree Program

as per
National Education Policy-2020

(Effective from the Session 2022-2023)

Head

Deptt. of Biotechnology
Faculty of Science
S.V.S.U., MEERUT
(Dr. Resmi Chaudhary)

SWAMI VIVEKANAND SUBHARTI UNIVERSITY
MEERUT



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(Dr. Raj Singh)

Dr. Ashwani Kumar

(Dr. Ashwani Kumar)

Dr. Manjul Muglani

(Dr. Manjul Muglani)

Semester-wise Titles of the Papers in B.Sc (Biochemistry)

Year	Sem	Course Code	Paper title	Theory/practical	credits
1.	I	B110101T	Fundamentals of Biochemistry	Theory	4
		B110102P	Biosafety Measures, Preparation of Solutions and Qualitative Analysis of Biomolecules	Practical/Field Work	2
	II	B110201T	Human Physiology and Clinical Biochemistry	Theory	4
		B110202P	Clinical Biochemistry Lab	Practical/Field Work	2
2.	III	B110301T	Tools and Techniques in Biochemistry	Theory	4
		B110302P	Biochemical Tools and Techniques Lab.	Practical	
	IV	B110401T	Enzymology and Immunology	Theory	4
		B110402P	Enzymes and Immunological Techniques Lab	Practical	2
3	V	B110501T	Bioenergetics and Metabolism	Theory	4
		B110502T	Fundamentals of Microbiology	Theory	4
		B110503P	Microbial Techniques and Metabolism Lab	Practical	2
	VI	B110601T	Cell, Molecular Biology and Genetic Engineering	Theory	4
		B110602T	Biostatistics, Bioinformatics and computer application in Biochemistry	Theory	4
		B110603P	Genetic Engineering and Bioinformatics Lab	Practical	2

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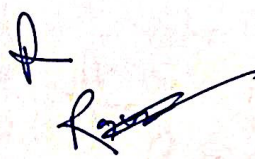
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Proposed Year wise Structure of UG Program in Biochemistry

Program/Year	Sem.	Course code	Paper title	Credits	Teaching hours
1 Certificate course in clinical biochemistry	1	B110101T	Fundamentals of Biochemistry	4	60
		B110102P	Biosafety Measures, Preparation of Solutions and Qualitative Analysis of Biomolecules	2	60
	2	B110201T	Human Physiology and Clinical Biochemistry	4	60
		B110202P	Clinical Biochemistry Lab	2	60
2 Diploma in tools and techniques in biochemistry	3	B110301T	Tools and Techniques in Biochemistry	4	60
		B110302P	Biochemical Tools and Techniques Lab	2	60
	4	B110401T	Enzymology and Immunology	4	60
		B110402P	Enzymes and Immunological Techniques Lab	2	60
3 Degree in Bachelor of Science Biochemistry	5	B110501T	Bioenergetics and Metabolism	4	60
		B110502T	Fundamentals of Microbiology	4	60
		B110503P	Microbial Techniques and Metabolism Lab	2	60
	6	B110601T	Cell, Molecular Biology and Genetic Engineering	4	60
		B110602T	Biostatistics, Bioinformatics and computer application in Biochemistry	4	60
		B110603P	Genetic Engineering and Bioinformatics Lab	2	60



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Subject prerequisite

To study BIOCHEMISTRY at undergraduate, a student must have Chemistry, Biology and /or Biotechnology in Class 12.

Programme Objectives (POs)

1. The programme intends to develop a strong theoretical and practical background in various domains of biochemistry.
2. The programme includes details of biomolecules, clinical biochemistry, tools and techniques, enzymes, immunology, cell biology, molecular biology, genetic engineering, biostatistics, and bioinformatics to make the living system more interesting human studies, which is the need of the hour.
3. The practical courses will equip the students with laboratory skills in biochemistry. Students will be able to design and conduct experiments, as well as to analyze and interpret scientific data.
4. The programme will provide students with the knowledge and skill base that would enable them to undertake further studies in biochemistry and related areas or in multidisciplinary areas that involve biochemistry and help develop a range of generic skills that are relevant in enhancing entrepreneurship skills among students.
5. The students will be exposed to a wide range of careers that combine biology, plants, and medicine.

Certificate Course in Clinical Biochemistry

B.Sc. I Programme Specific Outcomes (PSOs)

PSO1	This course introduces fundamentals of structure and function of biomolecules. Students will be able to develop an understanding of: the inter relationships within and between anatomical and physiological systems of the human body.
PSO1	The students will develop the understanding of basic concepts of clinical biochemistry, they would be able to relate clinical disorders with metabolic processes.
PSO1	The students will learn the basic principles of biochemistry relevant to possibilities of employment and research. Stress will be rigorous learning of lab practices like accurate preparation of solutions, and buffers. The course is intended to develop a sound, fundamental understanding of Biomolecular testing.
PSO1	The students will have hands-on training on qualitative estimation of important which will help them in getting employment in pathology labs and contribute to health care system.
PSO1	This Certificate course will enable students to apply for technical positions in government and private labs, academic and research institutes.

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Diploma in tools and techniques in biochemistry

B.Sc. II Programme based outcomes

PSO 1	Students will develop an understanding of: Principle, working, and applications of Biochemical tools & techniques to prepares them for independent execution of laboratory experiments using standard methods and techniques.
PSO 2	The objective of this course is to develop an understanding of the concepts of enzyme and enzyme kinetics.
PSO3	The students will develop an understanding of the basics of Immunology, types of Immune Responses, antigens and antibodies, histocompatibility, vaccines, and immunization. The students will develop a capability to function as paramedical staff during the current COVID crisis also.
PSO4	The course aims to develop an understanding of the concepts of enzyme dynamics. The students will also have understanding of basics of immunology, types of Blood grouping, cell counts, ELISA, Ouchterlony Double diffusion (ODD) and Separation of serum from blood & precipitation of Immunoglobulins
PSO5	The Diploma courses will ensure employability in Hospitals/Diagnostics and Pathology labs with good hands-on training. It will also enable students to take up higher studies and Research as their career and work in renowned national and international labs. Students can have their own start-ups as well.

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Degree in Bachelor of Science	
B.Sc III Programme Specific Outcomes (PSOs)	
PSO1	The student at the completion of the course will be able to have a detailed and conceptual understanding of molecular processes.
PSO2	The students will be able to understand and apply the principles and techniques of molecular biology which prepares students for further career in molecular biology. Independently execute a laboratory experiment using the standard methods and techniques.
PSO3	The principles of genetic engineering, gene cloning and related technologies will enable students to play an important role in applications of biotechnology in various fields like agriculture, forensic sciences, industry and human health and make a career out of it. Students can have their own start-ups as well.
PSO4	The basic tools of bioinformatics will enable students to analyze large amount of genomic data and its application to evolutionary biology. Apply knowledge and awareness of the basic principles and concepts of biology, computer science and mathematics existing software effectively to extract information from large databases and to use this information in computer modeling.
PSO5	The Degree courses will enable students to go for higher studies like Masters and Ph.D in Biochemistry and Allied subjects.

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Programme/Class: Certificate		Year: First	Semester: First
Subject: Biochemistry			
Course Code: B110101 T		Course Title: Fundamental of Biochemistry	
Course outcomes: The student at the completion of the course will learn to understand: <ul style="list-style-type: none"> • Basic details of structure, function of carbohydrate molecules and its classification • Details of structure, function and classification of amino acid & structural levels of protein molecules • Structure and function of fatty acids, storage and structural lipids • Details of structure and Function of Nucleotide, DNA and RNA • Basic details of Vitamin molecules and its classification • Classification, structural features and Function of Plant & Animal Hormone 			
Credits: 4		Core Compulsory	
Max. Marks: 25+75		Min. Passing Marks: As per rules	
Total No. of Lectures-Tutorials-Practical (in hours per week): L-T-P: 4-0-0			
Unit	Topics		No. of Lectures (60)
I	Basics of Biochemistry <ul style="list-style-type: none"> • History of biochemistry with special reference to contribution of Indian biochemists. • General idea about normality, molarity, molality, percentage solutions, mole fraction. W/v and v/v solutions. • Concept of pH determinations using indicators, buffer solutions and their biological importance. • Water as universal solvent 		5
II	Amino acids and proteins <ul style="list-style-type: none"> • Structural features and classification, Physical properties, optical properties (Stereoisomerism) • Chemical properties of amino acids • Uncommon amino acids and their function. • Classification of protein, structural organization as primary, secondary, tertiary and quaternary structure of protein and characteristics of the peptide bond 		10
III	Carbohydrate <ul style="list-style-type: none"> • Monosaccharides - structure of aldoses and ketoses, Ring structure of sugars, conformations of sugars, mutarotation, anomers, epimers and enantiomers • Structure of biologically important sugar derivatives, oxidation and reduction of sugars • Formation of disaccharides, reducing and non-reducing disaccharide • Polysaccharides - homo- and heteropolysaccharides, structural and storage polysaccharides 		10

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IV	Lipids <ul style="list-style-type: none"> • Building blocks of lipids - fatty acids, glycerol, ceramide • Storage lipids - triacyl glycerol and waxes • Structural lipids in membranes - glycerophospholipids, galactolipids and sulpholipids, sphingolipids and sterols • Plant steroids 	10
V	Nucleic acids <ul style="list-style-type: none"> • Nucleotides - structure and properties • Nucleic acid structure - Watson-Crick model of DNA • Structure of major species of RNA - mRNA, tRNA and rRNA • Nucleic acid chemistry - UV absorption, effect of acid and alkali on DNA • Other functions of nucleotides - source of energy, component of coenzymes, second messengers 	10
VI	Vitamins <ul style="list-style-type: none"> • Structure and active forms of water soluble and fatsoluble vitamins, • Deficiency diseases and symptoms, hypervitaminosis • Sources, dietary requirements 	5
VII	Plant Hormones <ul style="list-style-type: none"> • Classification, structural features & functions in Plants: • Auxins, gibberellins, Cytokinins, ethylene, and abscisic acid 	5
VIII	Animal Hormones <ul style="list-style-type: none"> • Classification, structural features & Functions of hormones secreted by endocrine glands: Hypothalamus, pituitary gland- anterior pituitary and posterior pituitary, thyroid gland, adrenal gland, Pancreas, gonads 	5

Suggested readings

1. Lehninger, Albert, Cox, Michael M., Nelson, David L. (2017) *Lehninger principles of biochemistry*/ New York: W.H. Freeman.
2. Voet, D., & Voet, J.G. (2011). *Biochemistry*. New York: J. Wiley & Sons
3. *Biochemistry - Lubert & Strayer* Freeman International Edition.
4. *Biochemistry - Keshav Trehan* Wiley Eastern Publications
5. *Fundamentals of Biochemistry - J.L. Jain* S. Chand and Company
6. Voet & Voet: *Biochemistry Vols 1 & 2*: Wiley (2004)
7. Murray et al: *Harper's Illustrated Biochemistry*: McGraw Hill (2003) Elliott and Elliott:
8. *Biochemistry and Molecular Biology*: Oxford University Press
9. Taiz, L., Zeiger, E., *Plant Physiology*. Sinauer Associates Inc., U.S.A. 5th Edition.
10. Hopkins, W.G., Huner, N.P., *Introduction to Plant Physiology*. John Wiley & Sons,
11. *Vander's Human Physiology* (2008) 11th ed., Widmaier, E.P., Raff, H. and Strang, K.T. McGraw Hill International Publications, ISBN: 978-0-07-128366-3.
12. *Endocrinology* (2007) 6th ed., Hadley, M.C. and Levine, J.E. Pearson Education (New Delhi), Inc. ISBN: 978-81-317-2610-5.

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Course Books published in Hindi must be prescribed by the Universities and Colleges

Course prerequisites: To study this course, a student must have had the subject Biology/Biotechnology/Chemistry in class/12th/ certificate/diploma.

Suggested Continuous Evaluation Methods:

Total Marks: 25

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

At the End of the whole syllabus any remarks/ suggestions: None

- Safety measures in laboratories
- Preparation of normal and molar solutions
- Preparation of buffers
- Determination of pKa of acetic acid and glycine
- Qualitative tests for carbohydrates, lipids, amino acids, proteins and nucleic acids
- Estimation of vitamin C
- Perform spot test for amino acids in a given sample

Suggested readings

1. Principles of Biochemistry- Albert L. Lehninger CBS Publishers & Distributors
2. Text-Book of Biochemistry with Clinical Correlations (2011) 7th ed., Devlin, T.M. John Wiley & Sons, Inc. (New York), ISBN-978-0-470-23177-4
3. An Introductory to practical Biochemistry, David T. Plummer (2006) Tata McGraw Hill Education, 3rd edition

Course books published in Hindi must be prescribed by the Universities and Colleges
Course prerequisites: To study this course, a student must have had the subject Biology/Biotechnology/Chemistry in class/12th/ certificate/diploma.
The digit, for this paper is 30 (from Arts/ Commerce/ Science)

Suggested Continuous Evaluation Methods:

Total Marks: 25

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

Programme/Class: Certificate	Year: First	Semester: First
Subject: Biochemistry		
Course Code: B110102 P	Course Title: Biosafety Measures, Preparation of Solutions and Qualitative Analysis of Biomolecules	
Course outcomes: After the successful course completion, learners will develop following attributes <ul style="list-style-type: none"> • Preparation of various solutions • Preparation of Buffers • Perform Qualitative test of Biomolecules • Estimation of vitamin C • Perform spot test for amino acids in a given sample 		
Credits: 4	Core Compulsory	
Max. Marks: 25+75	Min. Passing Marks: As per rules	
Total No. of Lectures-Tutorials-Practical (in hours per week): L-T-P: 0-0-4		
	Topics	Total No. of Lectures
I	<ul style="list-style-type: none"> • Safety measures in laboratories • Preparation of normal and molar solutions • Preparation of buffers • Determination of pKa of acetic acid and glycine • Qualitative tests for carbohydrates, lipids, amino acids, proteins and nucleic acids • Estimation of vitamin C • Perform spot test for amino acids in a given sample 	60
Suggested readings <ol style="list-style-type: none"> 1. Principles of Biochemistry- Albert L. Lehninger CBS Publishers & Distributors 2. Textbook of Biochemistry with Clinical Correlations (2011) 7th ed., Devlin, T.M., John Wiley & Sons, Inc. (New York), ISBN:978-0-470-28173-4. 3. An Introduction to Practical Biochemistry, David T. Plummer (2006) Tata McGraw Hill Education, 3rd edition 		
Course Books published in Hindi must be prescribed by the Universities and Colleges		
Course prerequisites: To study this course, a student must have had the subject Biology/Biotechnology/Chemistry in class/12 th / certificate/diploma. The eligibility for this paper is 10+2 from Arts/ Commerce/ Science		
Suggested Continuous Evaluation Methods:		
Total Marks: 25 House Examination/Test: 10 Marks Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks Class performance/Participation: 5 Marks		
Further Suggestions: None		

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Programme/Class: Certificate		Year: First	Semester: Second
Subject: Biochemistry			
Course Code: B110201 T		Course Title: Human Physiology and Clinical Biochemistry	
Course outcomes-			
After the successful course completion, learners will develop following attributes			
<ul style="list-style-type: none"> • Develop an understanding of the inter relationships within and between anatomical and physiological systems of the human body. • Develop the understanding of basic concepts of clinical biochemistry. • To understand disorder related with bio molecules metabolism. • Anticoagulant preservatives for blood and urine. • Metabolism of bilirubin, jaundice - types, differential diagnosis and Liver function. 			
Credits: 4		Core Compulsory	
Max. Marks: 25+75		Min. Passing Marks:	
Total No. of Lectures (in hours per week):			
Unit	Topics	No. of Lectures (60)	
I	Digestion and Respiration <ul style="list-style-type: none"> • Structural organization and functions of gastrointestinal tract and associated glands • Mechanical and chemical and enzymatic digestion of food, Absorptions of • carbohydrates, lipids, proteins, water, minerals and vitamins, • Mechanism of respiration, Pulmonary ventilation, Respiratory volumes and capacities, Transport of oxygen and carbon dioxide in blood Respiratory pigments, Dissociation curves and the factors influencing it, Control of respiration 	8	
II	Circulation and Excretion <ul style="list-style-type: none"> • Components of blood and their functions Haemostasis: Blood clotting system, Blood groups: Rh factor, ABO and MN • Cardiac cycle, Cardiac output and its regulation, Electrocardiogram, Blood pressure and its regulation • Structure of kidney and its functional unit, Mechanism of urine formation 	8	
III	Nervous System and Muscular System <ul style="list-style-type: none"> • Structure of neuron, and physiology of nerve impulse transmission • Histology of different types of muscle, Ultra structure of skeletal muscle • Molecular and chemical basis of muscle contraction • Control of muscle contraction by nerve impulses 	8	

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IV	Basic concepts of Clinical Biochemistry <ul style="list-style-type: none"> • A Brief review of units and abbreviations used in expressing concentrations and standard solutions • Specimen collection and processing (Blood, urine, feces) • Anticoagulant and preservatives for blood and urine samples • Transport of specimens 	8
V	Hematology: Blood <ul style="list-style-type: none"> • Composition and functions of various components, • Anemia:- classifications, erythrocyte indices • Blood coagulation system, Clotting time, Bleeding time, Prothrombin time, RBC count, WBC count, Platelet count, Differential count • determination of Hb, PCV and ESR. Hemoglobinopathies, Thalassemia 	8
VI	Disorders of Carbohydrate metabolism <ul style="list-style-type: none"> • Regulation of blood sugar • Glycosuria-types of Glycosuria • Oral glucose tolerance test in normal and diabetic condition • Diabetes mellitus and Diabetic insipidus - hypoglycemia, hyperglycemia. Ketonuria, ketosis 	4
VII	Disorders of Lipid metabolism <ul style="list-style-type: none"> • Cholesterol: Factors affecting blood cholesterol level • Dyslipoproteinemia, atherosclerosis risk factor and fatty liver. • Involvement of enzymes in diagnostics of heart disease including aspartate transaminase, isoenzymes of creatine kinase and lactate dehydrogenase and troponin 	4
VIII	Liver function test <ul style="list-style-type: none"> • Types, differential diagnosis • Liver function test - Icteric index, Vandenberg test, plasma protein changes. Renal function test: Clearance test-Urea, Creatinine • Para- aminohippuric acid (PAH) test, Concentration and dilution test. Enzymology: Clinical significance of SGOT, SGPT, ALP, ACP, CPK and LDH 	8

Suggested readings

1. Textbook of Medical Physiology by Guyton. A.C., H. Sanders Philadelphia. 1988.
 2. Physiological basis of Medical practice, West J.B., Best and Taylor.
 3. Introduction to Physiology by Davidson H and Segal M.B. Academic Press.
 4. Sherwood L - Human Pysiology: From Cells to Systems, (Wadsworth Publishing, 2000, ISBN: 0534568262)
 5. Tortora G J Principles of Anatomy & Physiology, (John Wiley & Sons, 1999, ISBN: 0471366927)
 6. Medical Biochemistry by MN Chatterjee, Rana Shinde, 8 edition, 2013, Jaypee publications.
 7. Textbook of Medical Laboratory Technology by Praful B. Godkar and Darshan P. Godkarth
 8. Medical Laboratory Technology by Ramniksood, 5 Edition, 1999, Jaypee publishers.
 9. Text book of Biochemistry with clinical correlation, Thomas M. Devlin, 3rd edition, A. JohnWiley-Liss Inc. Publication.
 10. Practical Clinical Biochemistry, Harold Varley, 4th edition, CBS Publication and Distributors, New Delhi.
- Course Books published in Hindi must be prescribed by the Universities and Colleges

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Course prerequisites: To study this course, a student must have had the subject Biology/Biotechnology/Chemistry in class/12th/ certificate/diploma.

Suggested Continuous Evaluation Methods:

Total Marks: 25

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

At the End of the whole syllabus any remarks/ suggestions: None

S.No.	Topic	Total No. of Lectures
	<ul style="list-style-type: none"> Qualitative and quantitative analysis of urine proteins, 8, 10-fold proteins, Cl, U₂ Qualitative analysis of abnormal constituents in urine: glucose, albumin, bile pigments, bile salts and acetone bodies. Experiments on blood: (a) Estimation of haemoglobin by cyanmethemoglobin method (b) Determination of A/G ratio in serum. Isolation and estimation of serum cholesterol Enzyme assays: alkaline phosphatase, SGOT, SGPT Estimation of haemoglobin using Sahli's haemoglobinometer Measurement of blood pressure using a sphygmomanometer Recording of blood glucose level by using glucometer Nephelometric test for N-acetyl amino acids Test for sugar and acetone in urine 	60
<p>Recommended Reading:</p> <ol style="list-style-type: none"> 1. Modi A.B., Biochemistry by M.D. Chatterjee, Rama Shukla, 8 edition, 2012, Jaypee publishers 2. Textbook of Medical Laboratory Technology by Preeti S. G. Jha and Geetika P. Chakrabarti 3. Myler, J. et al., Laboratory Investigation, 3 Edition, 1996, Jaypee publishers 4. Textbook of Biochemistry, 5th edition, Gnanaprakasam, Tripathi, M. Debnath, and others, A. Ghosh, Jaypee publishers 5. Practical Biochemistry, 4th edition, P. D. Das, Jaypee publishers and distributors, Jaypee 	<p>Handwritten notes and signatures: "A", "Rev", "2/3", "N"</p>	
<p>Total Marks: 25</p> <p>House Examination/Test: 10 Marks</p> <p>Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks</p> <p>Class performance/Participation: 5 Marks</p>		

Programme/Class: certificate	Year: First	Semester: Second
Subject: Biochemistry		
Course Code: B110201 T	Course Title: Clinical Biochemistry Lab	
Course outcomes-		
<ul style="list-style-type: none"> To learn qualitative and quantitative analysis of constituents of biological fluids such as urine, blood and their estimation using standard methods. Students will able to Perform basic hematological laboratory testing 		
Credits: 4	Core Compulsory	
Max. Marks: 25+75Min.	Passing Marks:As per rules	
Total No. of Lectures-Tutorials-Practical (in hours per week): L-T-P: 0-0-4		
UNIT	Topic	Total No.of Lectures
	<ul style="list-style-type: none"> Qualitative and quantitative analysis of urine : proteins, Bence-Jones proteins, Cl⁻, Ca⁺² Qualitative analysis of abnormal constituents in urine - glucose, albumin, bile pigments, bile salts and ketone bodies. Experiments on blood (a) Estimation of haemoglobin by cyanmethemoglobin method (b) Determination of A/G ratio in serum Isolation and estimation of serum cholesterol Serum enzyme assays: alkaline phosphatase, SGOT, SGPT Estimation of haemoglobin using Sahli's haemoglobinometer Recording of blood pressure using a sphygmomanometer Recording of blood glucose level by using glucometer Ninhydrin test for N-amino acids. Test for sugar and acetone in urine. 	60
Suggested Readings:		
<ol style="list-style-type: none"> Medical Biochemistry by MN Chatterjee, Rana Shinde, 8 edition, 2013, Jaypee publications. Textbook of Medical Laboratory Technology by Praful B. Godkar and Darshan P. Godkarth Medical Laboratory Technology by Ramniksood, 5 Edition, 1999, Jaypee publishers. Text book of Biochemistry with clinical correlation, Thomas M. Devlin, 3rd edition, A. JohnWiley-Liss Inc. Publication. Practical Clinical Biochemistry, Harold Varley, 4th edition, CBS Publication and Distributors, New Delhi. 		
Course Books published in Hindi must be prescribed by the Universities and Colleges		
Suggested Continuous Evaluation Methods:		
Total Marks: 25		
House Examination/Test: 10 Marks		
Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks		
Class performance/Participation: 5 Marks		
Further Suggestions: None		

At the End of the whole syllabus any remarks/ suggestions: None

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Programme/Class: DIPLOMA		Year: SECOND	Semester: THIRD
Subject: Biochemistry			
Course Code: B110301 T		Course Title: Tools and Technique in Biochemistry	
Course outcomes:			
<ul style="list-style-type: none"> The objective of the course is to introduce various techniques to the students, which are used in biological research. Students will acquire knowledge about the principles and applications of spectrophotometric and chromatography techniques used in a biochemistry lab. Students will learn about the principle and application of electrophoresis, centrifugation techniques, microscopic and molecular biological techniques. 			
Credits: 4		Core Compulsory	
Max. Marks: 25+75		Min. Passing Marks:	
Total No. of Lectures (in hours per week): L-T-P: 4-0-0			
Unit	Topics	No. of Lectures (60)	
I	Basics of Biophysics <ul style="list-style-type: none"> Chemical bonding - Ionic bond, covalent bond, hydrogen bond and Vander-Waals force. 	4	
II	Chromatography <ul style="list-style-type: none"> Introduction & Principle of Chromatography Paper, thin-layer, column, HPLC, GLC and molecular sieving., Ion exchange chromatography Affinity Chromatography 	8	
III	Centrifugation <ul style="list-style-type: none"> Principle of centrifugation Basic rules of sedimentation, sedimentation coefficient. Various types of centrifuges, low speed centrifuge, high speed centrifuge and ultracentrifuge, types of rotors. Application of centrifugation, differential centrifugation, density gradient centrifugation-zonal and isopycnic. 	8	
IV	Electrophoresis: <ul style="list-style-type: none"> Basic Principle of electrophoresis, Gel electrophoresis, PAGE, SDS-PAGE, Native gels, denaturing gels Agarose gel electrophoresis, 	8	
VI	Microscopy <ul style="list-style-type: none"> Principle of light microscopy, Phase contrast microscopy Fluorescence microscopy Electron microscopy Permanent and temporary slide preparation, histology and staining. 	8	
VII	Radioactivity <ul style="list-style-type: none"> Types, their importance in biological studies Measure of radioactivity GM counters and Scintillation counting. 	4	

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VIII	<p>Fundamental principles and basics of instrument design of:</p> <ul style="list-style-type: none"> • UV-Visible spectrophotometry and Beer-Lambert law • Fluorescence techniques • Infra-Red and Raman spectrometry • Circular Dichroism and Optical Rotatory dispersion • Nuclear Magnetic Resonance spectrometry • Atomic absorption and emission spectrometry • X Ray diffraction • Mass spectrometry 	8
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Suggested readings

1. Boyer, R.F., Biochemistry Laboratory: Modern Theory and Techniques, 6th ed., Boston, Mass: Prentice Hall, 2012,
 2. Plummer D. T., An Introduction to Practical Biochemistry 3rd ed., Tata McGraw Hill Education Pvt. Ltd. 2006.
 3. Wilson K. and Walker J., Principles and Techniques of Biochemistry and Molecular Biology, 7th ed., Cambridge University Press, 2010
 4. Rastogi & Pathak, Genetic Engineering, Oxford University Press, 2009
- Course Books published in Hindi must be prescribed by the Universities and Colleges.**

This course can be opted as an elective by the students of following subjects:

The eligibility for this paper is 10+2 with Biology/Biotechnology/Chemistry as one of the subject

Suggested Continuous Evaluation Methods:

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project/Research Orientation/ Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

At the End of the whole syllabus any remarks/ suggestions: None

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Programme/Class: DIPLOMA		Year: SECOND	Semester: THIRD
Course Code: B110201 T		Course Title: Biochemical Tools and Techniques Lab	
Course outcomes- It will also give them an opportunity to get hands on experience to develop their experimental skills expected from any biochemist working in a pathology/diagnostic/research lab.			
Credits: 4	Core Compulsory		
Max. Marks: 25+75Min.	Passing Marks: As per rules		
Total No. of Lectures-Tutorials-Practical (in hours per week): L-T-P: 0-0-4			
UNIT	Topic	Total No. of Lectures	
	<ul style="list-style-type: none"> • Verification of Beer's Law • Estimation of proteins by Biuret/Lowry method • Separation of amino acid acids by TLC/paper chromatography • To perform agarose gel electrophoresis • To isolate mitochondria by differential centrifugation • Visualization of cells by methylene blue • SDS PAGE 	60	
Suggested Readings:			
<ol style="list-style-type: none"> 1. Narayanan, P (2000) Essentials of Biophysics, New Age Int. Pub. New Delhi. 2. Roy R.N. (1999) A Text Book of Biophysics New Central Book Agency. 3. Plummer D. T., An Introduction to Practical Biochemistry 3rd ed., Tata McGraw Hill Education Pvt. Ltd. 1998, 4. Wilson K. and Walker J., Principles and Techniques of Biochemistry and Molecular Biology, 7th ed., Cambridge University Press, 2010 			
Course Books published in Hindi must be prescribed by the Universities and Colleges			
This course can be opted as an elective by the students of following subjects: The eligibility for this paper is 10+2 with Biology/Biotechnology/Chemistry as one of the subject			
Suggested Continuous Evaluation Methods:			
Total Marks: 25			
House Examination/Test: 10 Marks			
Written Assignment/Presentation/Project/ Research Orientation/ Term Papers/Seminar: 10 Marks			
Class performance/Participation: 5 Marks			
Further Suggestions: None			

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Programme/Class: DIPLOMA		Year: SECOND	Semester: FOURTH
Subject: Biochemistry			
Course Code: B110301 T		Course Title: Enzymes and Immunology	
Course outcomes:			
<ul style="list-style-type: none"> The objective of the course is to provide detailed knowledge about enzymes, the biological catalysts with remarkable properties that sustain life. Students will learn the nature and importance of enzymes in living systems Students will gain insight into the thermodynamic and molecular basis of catalysis by enzymes and the underlying basis of their specificity Students will learn about the mechanisms of enzyme action, kinetics of enzyme catalyzed reactions and clinical importance of enzyme inhibitors Students will also learn to appreciate how enzymes are regulated and the physiological importance of enzyme regulation in the cell Students will develop the understanding of basics of Immunology, types of Immune Responses, antigens and antibodies, histocompatibility, vaccines and immunization 			
Credits: 4		Core Compulsory	
Max. Marks: 25+75		Min. Passing Marks:	
Total No. of Lectures (in hours per week): L-T-P: 4-0-0			
Unit	Topics	No. of Lectures (60)	
I	Introduction to enzymes <ul style="list-style-type: none"> General characteristics of enzymes Co-factor and prosthetic group, apoenzyme, holoenzyme. Classification and nomenclature of enzymes. Enzyme assays- Enzyme activity, specific activity, units to express enzyme activity. Features of enzyme catalysis Catalytic power and specificity of enzymes (concept of active site), Fischer's lock and key hypothesis Koshland's induced fit hypothesis. 	8	
II	Enzyme kinetics <ul style="list-style-type: none"> Relationship between initial velocity and substrate concentration Michaelis-Menten equation Lineweaver-Burk plot, Eadie-Hofstee and Hanes plot Determination of Km and Vmax, Kcat, specificity constant Effect of pH and temperature on the activity of enzymes. 	8	
III	Enzyme inhibition and Regulation <ul style="list-style-type: none"> Reversible inhibition (competitive, uncompetitive, non-competitive and mixed) 	8	

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	<ul style="list-style-type: none"> • Irreversible inhibition • Substrate inhibition • Allosteric regulation and feedback inhibition (ATPase) • Isoenzymes • Enzyme immobilization and its applications 	
IV	<p>Introduction of Immunology</p> <ul style="list-style-type: none"> • Types of Immunity: Passive, Active, Innate and Acquired immunity, Humoral and Cell Mediated Immunity • Antigens: haptens, epitopes and Factors influencing immunogenicity • Antibodies: Structure, types, production and functions of immunoglobulins Clonal selection theory. • Antigen Antibody reaction: Precipitation, Immunoelectrophoresis, Haem-agglutination, RIA and ELISA. • Cell and organs of immune responses and their functions • B & T cells • factors responsible for immunogenicity • Monoclonal antibodies production and applications 	8
VI	<p>Histocompatibility</p> <ul style="list-style-type: none"> • Structure of MHC class I, II & III antigens and their mode of antigen presentation • MHC restriction, • Complement system: Components, Classical and alternate pathways of complement activation • Hypersensitivity • Autoimmunity. 	8
VII	<p>Vaccines and Immunization</p> <ul style="list-style-type: none"> • Passive and Active immunization • Types of Vaccines: Inactivated, Attenuated, Recombinant and Vaccines • Peptide and DNA Vaccines • RNA Vaccines 	4
VIII	<p>Transplantation immunology</p> <ul style="list-style-type: none"> • Immunological basis of graft rejection 	4

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- Clinical manifestations
- Immunosuppressive therapy and privileged sites

Suggested readings

1. Lehninger, AL "Principles of Biochemistry".
2. Lubert Stryer "Biochemistry".
3. Voet & Voet "Biochemistry".
4. Alan Fersht "Enzyme Structure and Mechanism".
5. David S. Sigman, Paul S. Sigman "The Enzymes: Mechanisms of Catalysis".
6. Trevor Palmer and Philip Bonner 2008 Enzymes Biochemistry, Biotechnology, Clinical Chemistry, 2nd edn EWP
7. Gerhartz W 2003 Enzymes in Industry Production and Applications, Wiley VCH
8. Wilson, K and Walker, J. (eds 2000 Principles and Techniques of Practical Biochemistry, 5th edn Cambridge University Press Palmer "Enzymes"
9. Dixon & Webb "Enzymes"
10. Kuby Immunology (2007) 6th ed., Kindt, T.L., Goldsby, R.A. and Osborne, B.A., W.H Freeman and Company (New York)
11. William, E. Paul (1989) Fundamental Immunology, 2nd Edition Raven Press, New York.
12. William, R. Clark (1991) the Experimental Foundations of Modern Immunology (4th Edition) John Wiley and Sons, New York.
13. Basic Immunology, A.K. Abbas and A.H. Lichtman, Saunders W.B. Company
14. Fundamentals of Immunology, W. Paul, Lippincott Williams and Wilkins
15. Immunology, W.L. Anderson, Fence Creek Publishing (Blackwell).

Course Books published in Hindi must be prescribed by the Universities and Colleges.

This course can be opted as an elective by the students of following subjects:

The eligibility for this paper is 10+2 with Biology/Biotechnology/Chemistry as one of the subject.

Suggested Continuous Evaluation Methods:

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project/Research Orientation / Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

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Programme/Class: DIPLOMA		Year: SECOND	Semester: FOURTH
Course Code: B110402P T		Course Title: Enzymes and Immunological Techniques Lab	
Course outcomes- After the successful course completion, learners will develop following attributes: <ul style="list-style-type: none"> • Know how to isolate enzyme and determine enzyme activity. • Know how to study the effect of pH and temperature on the enzyme activity. • Know how to study the effect of varying substrate and inhibitor concentration on the enzyme activity • Know how to detect Amino acids by Paper chromatography and TLC • This course aims to develop the understanding of basics of immunology, types of Blood grouping, cell counts, ELISA, Ouchterlony Double diffusion (ODD) and Separation of serum from blood & precipitation of Immunoglobulins • It will also give them an opportunity to get hands on experience to develop their experimental skills expected from any biochemist working in a pathology/diagnostic/research lab. 			
Credits: 4	Core Compulsory		
Max. Marks: 25+75Min.	Passing Marks: As per rules		
Total No. of Lectures-Tutorials-Practical (in hours per week): L-T-P: 0-0-4			
UNIT	Topic	Total No.of Lectures	
	<ul style="list-style-type: none"> • Isolation of enzyme and determination of enzyme activity • Study of the effect of pH on the enzyme activity. • Study of the effect of varying substrate concentration on the enzyme activity and determination of Km and Vmax. • Study of the effect of temperature on the enzyme activity. • Study of the effect of inhibitors on the enzyme activity. • Blood grouping • Differential Count of WBC • Detergent lysis of RBC • Dot ELISA • ELISA - Demonstration • Ouchterlony Double diffusion (ODD) • Separation of serum from blood & precipitation of Immunoglobulins 	60	
Suggested Readings: <ol style="list-style-type: none"> 1. Clark & Switzer. Experimental Biochemistry. Freeman (2000) 2. Trevor Palmer and Philip Bonner 2008 Enzymes Biochemistry, Biotechnology, Clinical Chemistry, 2nd edn EWP 3. Wilson, K and Walker, J ..(eds 2000 Principles and Techniques of Practical Biochemistry, 5th edn Cambridge University Press 4. Kuby Immunology (2007) 6th ed., Kindt, T.L., Goldsby, R.A. and Osborne, B.A., W.H Freeman and Company (New York) 5. William, E. Paul (1989) Fundamental Immunology, 2nd Edition Raven Press, New York. 6. William, R. Clark (1991) the Experimental Foundations of Modern Immunology (4th Edition) John Wiley and Sons, New York. 7. Basic Immunology, A.K. Abbas and A.H. Lichtman, Saunders W.B. Company 			
Course Books published in Hindi must be prescribed by the Universities and Colleges			
This course can be opted as an elective by the students of following subjects: The eligibility for this paper is 10+2 with Biology/Biotechnology/Chemistry as one of the subject			
Suggested Continuous Evaluation Methods: Total Marks: 25 House Examination/Test: 10 Marks			

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Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks
Class performance/Participation: 5 Marks

Further Suggestions: None

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Programme/Class: DEGREE	Year: THIRD	Semester: FIFTH
Subject: Biochemistry		
Course Code: B110501 T	Course Title Bioenergetics and Metabolism	
Course outcomes: The learners will be able to: <ul style="list-style-type: none"> • Understand the concepts of metabolism, characteristics of metabolic pathways and strategies used to study these pathways. • Gain a detailed knowledge of various catabolic and anabolic pathways • Understand the regulation of various pathways • Gain knowledge about the diseases caused by defects in metabolism with emphasis on the metabolic control 		
Credits: 4		Core Compulsory
Max. Marks: 25+75		Min. Passing Marks:
Total No. of Lectures (in hours per week):		
Unit	Topics	No. of Lectures
I	Principle of Bioenergetics: <ul style="list-style-type: none"> • Bioenergetics and thermodynamics, • Laws of Thermodynamics • Gibbs free energy, enthalpy • Entropy and their relationships • Free energy change • ATP as universal currency in biological system • Coenzymes and proteins as universal electron carriers 	60
II	Oxidative phosphorylation <ul style="list-style-type: none"> • The electron transport chain - its organization and function • Peter Mitchell's chemiosmotic hypothesis and Proton motive force • FoF₁ATP synthase, structure and mechanism of ATP synthesis • Metabolite transporters in mitochondria • Regulation of oxidative phosphorylation • ROS production and antioxidant mechanisms • Oxidative phosphorylation and ATP synthesis uncouplers 	4
III	Carbohydrate Metabolism: <ul style="list-style-type: none"> • Glycolysis • TCA cycle • Electron Transport Chain • Pentose phosphate pathway • Gluconeogenesis and Glycogen metabolism • Diseases associated with metabolic irregularities. 	8

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IV	Photosynthesis <ul style="list-style-type: none"> • Light harvesting and photosynthetic electron transport • Water splitting, formation of H⁺ gradient and photophosphorylation • Calvin cycle, and its regulation • Photo respiration • C4 and CAM pathways in plants 	8
V	Lipid Metabolism: <ul style="list-style-type: none"> • Degradation of fatty acids • β oxidation • regulation of fatty acid oxidation • ω oxidation and α oxidation • Ketone-body metabolism • Cholesterol synthesis • Fatty acid synthase complex enzyme • Synthesis of saturated, unsaturated, odd and even chain fatty acids • Regulation of fatty acid metabolism • Diseases associated with abnormal lipid metabolism 	8
VI	Protein Metabolism <ul style="list-style-type: none"> • Urea Cycle • Transport of ammonia • Deamination and transamination reactions • Inborn errors of protein metabolism • Glucogenic and ketogenic amino acids • Overview of amino acid synthesis 	8
VII	Nucleic Acid Metabolism <ul style="list-style-type: none"> • De novo synthesis of purine and pyrimidine nucleotides • regulation and salvage pathways • degradation of purine and pyrimidine nucleotides • Inhibitors of nucleotide metabolism • Disorders of purine and pyrimidine metabolism 	8
VIII	Nitrogen metabolism <ul style="list-style-type: none"> • Biological nitrogen fixation by free living and in symbiotic association Structure and function of the enzyme nitrogenase • Nitrate assimilation: Nitrate and Nitrite reductase • Primary and secondary ammonia assimilation in plants • ammonia assimilation by glutamine synthetase-glutamine oxoglutarate amino transferase (GS-GOGAT) pathway 	8

Suggested readings

1. Lehninger, Albert, Cox, Michael M., Nelson, David L. (2017) *Lehninger principles of biochemistry/*

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New York: W.H. Freeman.

2. Voet, D., & Voet, J.G. (2011). Biochemistry. New York: J. Wiley & Sons
3. Biochemistry - Lubertstryer Freeman International Edition.
4. Biochemistry - Keshav Trehan Wiley Eastern Publications
5. Fundamentals of Biochemistry - J.L. Jain S. Chand and Company
6. Voet & Voet: Biochemistry Vols 1 & 2: Wiley (2004)
7. Murray et al: Harper's Illustrated Biochemistry: McGraw Hill (2003) Elliott and Elliott:
8. Biochemistry and Molecular Biology: Oxford University Press
9. Taiz, L., Zeiger, E., Plant Physiology. Sinauer Associates Inc., U.S.A. 5th Edition.
10. Hopkins, W.G., Huner, N.P., Introduction to Plant Physiology. John Wiley & Sons,

This course can be opted as an elective by the students of following subjects:

The eligibility for this paper is 10+2 with Biology/Biotechnology/Chemistry as one of the subject

Suggested Continuous Evaluation Methods:

Total Marks: 25

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project/Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

III. Microbiology

- Contributions in Anton van Leeuwenhoek, Louis Pasteur, Robert Koch, Alexander Fleming
- Various forms of microorganisms (bacteria, fungi, viruses, protozoa, Pflanz)

Classification of microbiology

- Nutritional classification of microorganisms
- Morphology of microorganisms
- Chemotaxonomy and chemotaxonomic techniques
- Growth curves

Control of Microorganisms

- Physical agents (Heat, Cold, Radiation, Ultrasound and High Pressure)
- Chemical agents (Alcohol, Hydrogen peroxide, Chlorine, Iodine, Phenol, Formalin, Methicillin)

III. Microbiology: Microorganisms and Environmental Microbiology

- Microorganisms as pathogens (bacteria, fungi, viruses, protozoa and fungi)

IV. Microbiology: Microorganisms and Environmental Microbiology

- Determination of the level of microbial activity
- Differentiability test and disc diffusion test
- Range of activity and membership of various microorganisms

Microbes in various environments and microbial interactions

- The thermophiles and psychrophiles
- Symbiosis and endosymbiosis among microbial populations
- N₂ fixing microbes in agriculture and industry

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Programme/Class: DEGREE		Year: THIRD	Semester: FIFTH
Subject: Biochemistry			
Course Code: B110502 T		Course Title Fundamentals of Microbiology	
Course outcomes: After the successful course completion, learners will develop following attributes <ul style="list-style-type: none"> • Know the basics of microbiology • Have knowledge of the general classification of microbes • understand basics of Control of Microorganisms • Study microbes in extreme environments and microbial interactions • Know the basics of recombination in Prokaryotes • Food & Industrial Microbiology • Basics of virology 			
Credits: 4		Core Compulsory	
Max. Marks: 25+75		Min. Passing Marks:	
Total No. of Lectures (in hours per week):			
Unit	Topics		No. of Lectures (60)
I	History of Microbiology <ul style="list-style-type: none"> • Spontaneous generation versus biogenesis • Contributions of Anton von Leeuwenhoek, Louis Pasteur, Robert Koch, Alexander Fleming • Various forms of microorganisms (bacteria, fungi, viruses, protozoa, PPLOs) 		4
II	Classification of microbiology <ul style="list-style-type: none"> • Nutritional classification of microorganisms • Nature of the microbial cell surface • Gram positive and Gram negative bacteria • Growth curve 		8
III	Control of Microorganisms <ul style="list-style-type: none"> • Physical agents (Autoclave, Hot air oven, Laminar airflow and membrane filter.) • Chemical agents (Alcohol, Halogens and Gaseous agents antibiotics), Radiation Methods (UV rays) 		8
IV	Pathogenicity of Microorganisms and Antimicrobial Chemotherapy <ul style="list-style-type: none"> • Introduction to pathogenic microbes, Bacteria, Viruses, Algae, protozoa and fungi • General Characteristics of antimicrobial drugs • determining the level of microbial activity • dilution susceptibility test and disc diffusion test • Range of activity and mechanism of action of penicillin, vancomycin and tetracycline. 		8
V	Microbes in extreme environments and microbial interactions <ul style="list-style-type: none"> • The thermophiles alkalophiles, acidophiles • symbiosis and antibiosis among microbial population • N₂ fixing microbes in agriculture and forestry. 		8

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VI	Recombination in Prokaryotes <ul style="list-style-type: none"> • Transformation • Conjugation • Transduction 	4
VII	Food and Industrial Microbiology <ul style="list-style-type: none"> • Importance of microbiology in food and industries • Basic design of fermenter • Continuous and discontinuous culture • Preparation of fermented food products such as yoghurt, curd and cheese. Preparation of alcoholic beverages like wine and beer • Single cell proteins • Treatment of wastewater and sewage Bioremediation and biodegradation 	8
VIII	Brief outline of virology <ul style="list-style-type: none"> • Discovery of virus • Early development of virology • nomenclature • classification and taxonomy of viruses - based on host, nucleic acids and structure • Evolution of viruses 	8

Suggested readings

- Brock Biology of Microorganisms 11th edition and Brock Biology of Microorganisms ILLUSTRATIONS ISBN 0-13-196893-9 © Prentice Hall
- **MICROBIOLOGY - AN INTRODUCTION**, 8th edition Gerard J. Tortora, Bergen Community College by Berdell R. Funke, North Dakota State University Christine L. Case, Skyline College ©2004 | Pearson
- J. Willey, L. Sherwood & C. Woolverton, Prescott's Microbiology, 10th Ed., McGraw Hill international, (2017). ISBN 13: 9781259657573 2. MJ Chan, ECS Krieg & NR Pelczar, Microbiology, 5th Ed. McGraw Hill International, (2004)

Course prerequisites: To study this course, a student must have had the subject Biology/Biotechnology in class/12th/ certificate/diploma.

Suggested Continuous Evaluation Methods:

Total Marks: 25

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project / Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

At the End of the whole syllabus any remarks/ suggestions: None

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Programme/Class: DIGREE		Year: THIRD	Semester: FIFTH
Course Code: B110503P T	Course Title: Microbial Techniques and Metabolism Lab		
Course outcomes <ul style="list-style-type: none"> On successful completion of this paper, students should be able to: Perform enzyme assay Identify different microbes Perform routine microbiological practices including sterilization, media preparation, maintenance of microbial culture, staining etc. To carry out research using microbes. To test microbial culture for antibiotic resistance. 			
Credits: 4	Core Compulsory		
Max. Marks: 25+75 Min.	Passing Marks: As per rules		
Total No. of Lectures-Tutorials-Practical (in hours per week): L-T-P: 0-0-4			
UNIT	Topic	Total No. of Lectures	
	<ul style="list-style-type: none"> Enzyme assay (one example) Biochemical tests-starch hydrolysis, gelatin liquefaction. Assay of salivary amylase. Cholesterol estimation. Cleaning and sterilization of glassware. Study of instruments: Compound microscope, Autoclave, Hot air oven, pH meter, Laminar airflow and centrifuge Media preparation: Nutrients agar, Nutrient broth and LB. Staining Techniques: Simple, Negative staining, Gram staining, Endospore staining, fungal staining. Isolation of bacteria and fungi from soil/air/water - dilution and pour plate methods Study of Rhizobium from root nodules of legumes Growth curve of bacteria 	60	
Suggested Readings: <ul style="list-style-type: none"> Wilson, K and Walker, J ..(eds 2000 Principles and Techniques of Practical Biochemistry, 5th edn Cambridge University Press M.T. Madigan, J.M. Martinko & D.A. Stahl, Brock Biology of Microorganisms, 13th Ed., Pearson Education International. (2010) J.G. Cappuccino, and N. Sherman, Microbiology: A Laboratory manual, 10th Ed. Benjamin/Cummings (2013) 			
Course Books published in Hindi must be prescribed by the Universities and Colleges			
This course can be opted as an elective by the students of following subjects: The eligibility for this paper is 10+2 with Biology/Biotechnology/Chemistry as one of the subject			
Suggested Continuous Evaluation Methods: Total Marks: 25 House Examination/Test: 10 Marks Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks Class performance/Participation: 5 Marks			
Further Suggestions: None			

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Programme/Class: DEGREE	Year: THIRD	Semester: SIXTH
Subject: Biochemistry		
Course Code: B110601 T	Course Title: Cell, Molecular Biology and Genetic Engineering	
Course outcomes: After the successful course completion, learners will develop following attributes:		
<ul style="list-style-type: none"> • Distinguish between the cellular organization of prokaryotic and eukaryotic cells • Would have deeper understanding of cell at structural and functional level. • Will able to understand details of central dogma of life • Get proper knowledge about the DNA manipulative enzymes: Restriction enzymes and DNA ligases, and Gene cloning vectors. • Gain knowledge about In vitro construction of recombinant DNA molecules vector DNA • learn about screening and selection of recombinant host cells, Gene Libraries, cloning techniques, Expression of cloned DNA • Have knowledge of Application of r-DNA technique in human health and quality crop production 		
Credits: 4		Core Compulsory
Max. Marks: 25+75		Min. Passing Marks:
Total No. of Lectures (in hours per week):		
Unit	Topics	Total no. of Lectures (60)
I	Cell Biology: <ul style="list-style-type: none"> • Intracellular organization: • Cell Membrane, Fluid Mosaic Model, and membrane transport. • Structure and functions of organelles, • Prokaryotic and eukaryotic cell wall, • Cell cycle, cell death and cell renewal: • Eukaryotic cell cycle, restriction point, and checkpoints. • Cell division: Mitosis and Meiosis. • Apoptosis and necrosis 	4
II	<ul style="list-style-type: none"> • Fundamental principles of cell signalling. Concept of signalling as a two-box system • G-Protein and Receptor Tyrosine Kinase mediated signalling • Elements of eukaryotic cytoskeleton. Organisation and dynamics of actin microfilaments and microtubules • Endomembrane system, secretory pathways and vesicular trafficking 	8

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III	Basics of Molecular Biology: <ul style="list-style-type: none"> • Central dogma of Life • Organization of Genetic Material, • DNA Replication • Prokaryotic- Enzymes and proteins involved in replication • Spontaneous and induced mutations, • Physical and chemical mutagens, • Mutation at the molecular level, • DNA damage & Repair • Mutations in plants, animals, and microbes for economic benefit of man. 	10
IV	Transcription: <ul style="list-style-type: none"> • Transcription in prokaryotes, • Mechanism, Promoters • RNA polymerase • Transcription factors 	8
V	Translation: <ul style="list-style-type: none"> • Genetic code, • Properties and Wobble hypothesis. • Translation: Mechanism of translation in Prokaryotes • Regulation of Gene expression: • Regulation of Gene expression in Prokaryotes: • Operon concept (Lac) 	8
VI	Recombinant DNA Technology: <ul style="list-style-type: none"> • DNA manipulative enzymes • Restriction enzymes and DNA ligases, • Gene cloning vectors: Plasmids, Bacteriophage and Chimeric plasmids, • Creation of r-DNA, • Transformation of r-DNA by different methods, • Screening and selection of recombinant host cells, • Gene Libraries: Genomic DNA and cDNA cloning techniques 	8
VII	Applications of r-DNA technique in human health <ul style="list-style-type: none"> • Production of Insulin, • Production of recombinant vaccines: Hepatitis B, • Production of human growth hormone 	6

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VIII	Transgenic plants <ul style="list-style-type: none"> • Methods of plant transformation • Agrobacterium mediated plant transformation • Application of plant genetic engineering: • Insect resistance, • Disease resistance, • Herbicide resistance • Abiotic stress tolerance • Delayed fruit ripening 	8
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Suggested readings

1. Lehninger, Albert L., Cox, Michael M., Nelson, David L. (2017) *Lehninger principles of biochemistry* / New York : W.H. Freeman
2. Watson, J. D., Baker, T. A., Bell, S. P., Gann, A., Levine, M., & Losick, R. M. (2013). **Molecular biology of the gene.**
3. Voet, D., & Voet, J. G. (2011). *Biochemistry*. New York: J. Wiley & Sons.
4. Ulrich Hubscher, Giovanni Maga, and Silvio Spadari (2007), *Eukaryotic dna polymerases* *Annu. Rev. Biochem.* 2002. 71:133-63
DOI:10.1146/annurev.biochem.71.090501.150041.
5. Smita Rastogi and Neelam Pathak (2009), *Genetic Engineering*, Oxford University Press.
6. *Gene Cloning and DNA Analysis* (2010) 6th ed., Brown, T.A., Wiley-Blackwell publishing (Oxford, *Principles of Gene Manipulation and Genomics* (2006) 7th ed., Primrose, S.B., and Twyman, R. M., Blackwell publishing (Oxford, UK)
7. *Molecular Biotechnology: Principles and Applications of Recombinant DNA* (2010) 4th ed., Glick B.R., Pasternak, J.J. and Patten, C.L., ASM Press (Washington DC),
8. *Molecular Cloning: A laboratory manual* (2014), 4th ed., Michael R Green and J. Sambrook Cold spring Harbor laboratory press (3vol.), ISBN: 978-1-936113-42-2

Course prerequisites: To study this course, a student must have had the subject **Biology/Biotechnology/Chemistry** in class/12th/certificate/diploma.

Suggested Continuous Evaluation Methods:

Total Marks: 25

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project/Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

At the End of the whole syllabus any remarks/ suggestions: None

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Programme/Class: DEGREE	Year: THIRD	Semester: SIXTH
Subject: Biochemistry		
Course Code: B110602 T	Course Title: Biostatistics, Bioinformatics and computer application in Biochemistry	
Course outcomes: After the successful course completion, learners will develop following attributes: <ul style="list-style-type: none"> • Understand the principles of biological data collection, statistical analysis and presentation. • Learn and appreciate various factors that influence type of sample collected and sample size. • Collect, analyze and interpret biological data using appropriate statistical tools • Improve their computational, mathematical and computer skills, which would increase their eligibility to pursue research based higher education. • Formulate and justify appropriate choices in technology, strategy, and analysis for a range of projects involving DNA, RNA, or protein sequence data. • Explain common methods and applications for analysis of gene or protein expression. • Use data visualization software to effectively communicate results. 		
Credits: 4		Core Compulsory
Max. Marks: 25+75		Min. Passing Marks:
Total No. of Lectures (in hours per week):		
Unit	Topics	No. of Lectures (60)
I	Handling of data <ul style="list-style-type: none"> • Tabulation and diagrammatic representation of data • Bar diagram and pie diagram. • Measures of central tendency: mean, median and mode. • Measures of dispersion: range, quartile deviation, mean deviation and standard deviation. • Coefficient of variation. 	4
II	Tests of significance: <ul style="list-style-type: none"> • Null hypothesis and alternative hypothesis, • Z-test, • Student's distribution, • Paired t - test, • F-test for equality of population variances. • Contingency table, • Chi-square test for goodness of fit and independence of attributes, Correlation analysis 	8
III	<ul style="list-style-type: none"> • Molecular Techniques • DNA sequencing, Polymerase Chain Reaction (PCR) • Primer designing, DNA fingerprinting, site directed mutagenesis, RFLP, RAPD • Southern, Northern and Western Blotting 	4

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IV	Basics of Computer and Bioinformatics <ul style="list-style-type: none"> • Operating systems • Hardware, Software, • DOS, Data Access Using Data Control • Internet, LAN, WAN, Web servers. • MS word office, excel ,powerpoint • Definition and need of Bioinformatics, • Brief history of biological databases • International nucleotide databases (e.g., Gen Bank, European Molecular Biology Laboratory (EMBL) • Bio information and DNA Data Bank of Japan (DDBJ) Center) • International Nucleotide Sequence Database Collaboration (INSDC). 	8
V	Protein Databases <ul style="list-style-type: none"> • Classification of protein databases (e.g., primary, secondary, and composite databases) • Brief overview of ExPASy (Expert Protein Analysis System) bioinformatics resource portal • Protein 3D structural databases (e.g., RCSB-PDB (Research Collaboratory for Structural Bioinformatics Protein Data Bank), and MMDB (Molecular Modeling Database) of NCBI) 	8
	Database Similarity Searches: <ul style="list-style-type: none"> • BLAST, • FASTA, • PSI-BLAST, algorithms, • Multiple sequence alignments - CLUSTAL, PRAS. Primer Designing, • Homology Modeling, • Phylogenetic analysis • Drug Designing, • Determination of Secondary & Tertiary of proteins. 	8
VII	Biological File Formats and Literatures Databases <ul style="list-style-type: none"> • Brief overview of biological sequence and 3D structure file formats (e.g., GenBank/GenPept, EMBL, FASTA, PIR, and PDB), • NCBI's literature databases (e.g., PubMed, PubMed Central, PubChem Project and OMIM database 	8
VIII	Database Similarity Searching and Phylogenetics <ul style="list-style-type: none"> • Requirements of database searching, • BLAST (Basic Local Alignment Search Tool) algorithm, • Statistical significance and variants of BLAST • FASTA algorithm and its statistical significance • Comparison of BLAST and FASTA • Brief Overview of phylogenetic analysis 	8
Suggested readings <ol style="list-style-type: none"> 1. Analysis of biological data, M. Whitlock and D. Schluter (2009), Roberts and company publishers 2. Principles of biostatistics, M. Pagano and K. Gauvreau (2000), Duxbury Thomas learnings 		

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3. Protein Bioinformatics: From Sequence to Function, Academic Press, 2011, ISBN 0123884241, 9780123884244.
4. Essential Bioinformatics, Cambridge University Press, 2006, ISBN 113945062X, 9781139450621
5. Kerns EH, Di L. Drug-Like Properties: Concepts, Structure Design and Methods: from ADME to Toxicity Optimization, Academic Press, Oxford, 2008

Course prerequisites: To study this course, a student must have had the subject Biology/Biotechnology/Chemistry in class/12th/certificate/diploma.

Suggested Continuous Evaluation Methods:

Total Marks: 25

House Examination/Test: 10 Marks

Written Assignment/Presentation/Project/Term Papers/Seminar: 10 Marks

Class performance/Participation: 5 Marks

Further Suggestions: None

At the End of the whole syllabus any remarks/ suggestions: None

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Programme/Class: DIGREE	Year: THIRD	Semester: Sixth
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Course Code: B110603P T	Course Title: Genetic Engineering and Bioinformatics Lab
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Course outcomes
 On completion of this course, students will be able to:

- Isolate genomic DNA from bacteria, plant and animal tissues
- Isolate plasmid DNA (E. coli)
- Perform restriction digestion of DNA
- Perform Agarose Gel Electrophoresis
- Develop understanding of Bioinformatics as tools for Sequence Alignment, FASTA & BLAST search, Multiple Sequence Alignment, Protein Structure Visualization, as well as for Gene Finding

Credits: 4	Core Compulsory
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Max. Marks: 25+75 Min.	Passing Marks: As per rules
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Total No. of Lectures-Tutorials-Practical (in hours per week): L-T-P: 0-0-4

UNIT	Topic	Total No. of Lectures
	<ul style="list-style-type: none"> • Isolate genomic DNA from bacteria, plant and animal tissues • Isolate plasmid DNA (E. coli) • Perform restriction digestion of DNA • Perform Agarose Gel Electrophoresis • Learning to analyze data using SPSS or R software • Introduction to types of sequence databases (Nucleotides & Protein) • Pair wise Sequence Alignment (NW and SW approach) • FASTA & BLAST search • Multiple Sequence Alignment (ClustalX&Treeview) 	60

Suggested Readings:

1. Molecular Cloning: A laboratory manual (2014), 4th ed., Michael R Green and J. Sambrook Cold spring Harbor laboratory press (3vol.),
2. Bioinformatics - Principles and Applications (2008), 1st ed. Ghosh, Z. and Mallick, B., Oxford University Press (India)

Course Books published in Hindi must be prescribed by the Universities and Colleges

This course can be opted as an elective by the students of following subjects:
 The eligibility for this paper is 10+2 with Biology/Biotechnology/Chemistry as one of the subject

Suggested Continuous Evaluation Methods:
 Total Marks: 25
 House Examination/Test: 10 Marks
 Written Assignment/Presentation/Project / Research Orientation/ Term Papers/Seminar: 10 Marks
 Class performance/Participation: 5 Marks

Further Suggestions: None

Head
 Deptt. of Biotechnology
 Faculty of Science
 S.V.S.U., MEERUT



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